

# Mycoplasma Gene Detection Kit Myco Finder (Product Code: 69202)

## Operation Manual

---

# Kit Composition (Detection Reagent)

For sales rep training

## Mycoplasma Gene Detection Kit, Myco Finder (48 tests)

### [Appearance]



### [Constituent reagents]

- |   |   |
|---|---|
| (1) Test strip A (white)                                | 6 |
| (2) Test strip B (transparent)                          | 6 |
| (3) Positive control ( $2 \times 10^3$ copies/ $\mu$ L) | 1 |
| (4) Negative control (DNase Free Water)                 | 1 |
| (5) Flat cap  | 6 |



- Micro pipet (Several types: so that 5~1000  $\mu$ L can be measured to be taken)
- Filter tip (Sterilized, DNase · RNase Free)
- Mixer
- Heat block (Water Bath can also be used)
- Centrifuge (Capable of maximum speed 20,000 x g)
- 1.5 mL Micro tube
- Real-time PCR system (Capable of detecting FAM + HEX or ROX)

Micro pipet



Heat block



Real-time PCR



Centrifuge

14,000 rpm is required



1.5 mL Micro tube



Mixer

# Reagents for DNA Extraction

For sales rep training

DNA extraction is performed before using the mycoplasma gene detection kit.

## ● Introduction of QIAGEN QIAamp UCP DNA Micro Kit Reagents to be used:

**A : Proteinase K**

**B : Buffer AUL**

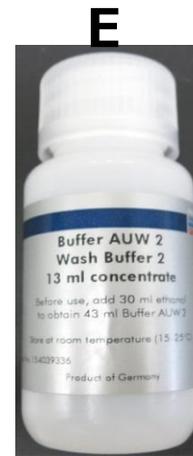
**C : QIAamp UCP MinElute Column**

**D : Buffer AUW1**

**E : Buffer AUW2**

**F : Buffer AUE**

**G : Collection Tube**



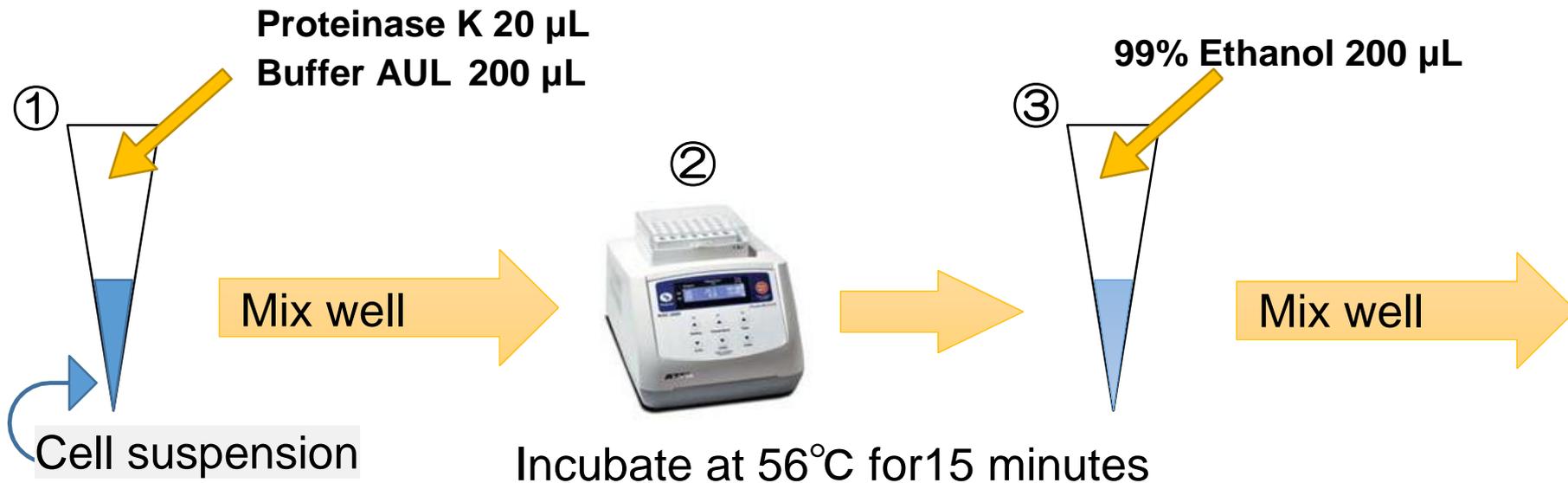
**Note) Certain reagents need to be prepared before use. Please follow the instructions on the respective bottle.**

# DNA Extraction Procedure (1)

For sales rep training

## [Extraction using QIAamp UCP DNA Micro Kit (1)]

- 1) Add **20 $\mu$ L** of **Proteinase K** and **200 $\mu$ L** of **Buffer AL** to a 200 $\mu$ L of cell culture or a **200 $\mu$ L** of **cell suspension** prepared by concentrating cell culture by centrifugation, and mix well by a mixer.  
Note) Mix well until the solution is homogenized.
- 2) Incubate at 56 $^{\circ}$ C for 15 minutes. After incubation, spin down to get rid of the liquid on the lid
- 3) Add **200 $\mu$ L** of **99 % ethanol** and mix well by a mixer.  
Note) Mix well until the solution is homogenized.

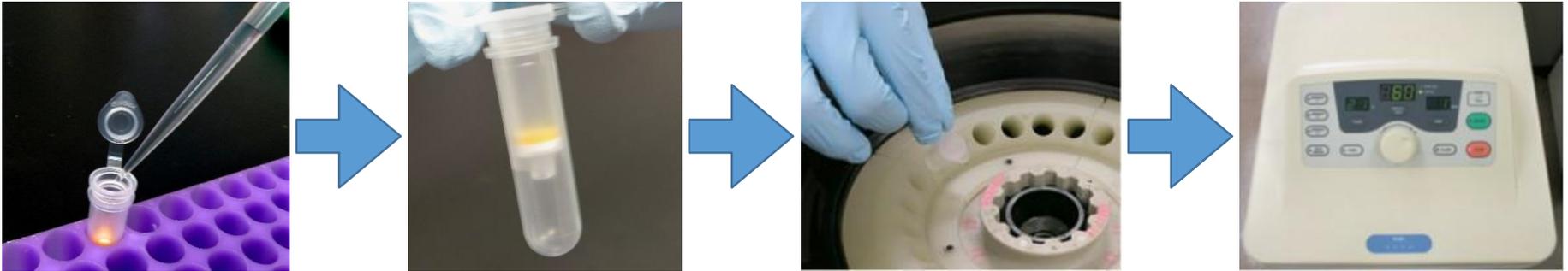


# DNA Extraction Procedure (2)

For sales rep training

[Extraction using QIAamp UCP DNA Micro Kit (2)]

4) Move all into **QIAamp UCP MinElute Column** and centrifuge at  $6,000 \times g$  (8,000 rpm) for 1 minute.



5) Place the **QIAamp UCP MinElute Column** into a new collection tube and add **500 $\mu$ L** of **Buffer AW1**. Centrifuge at  $6,000 \times g$  (8,000 rpm) for 1 minute.

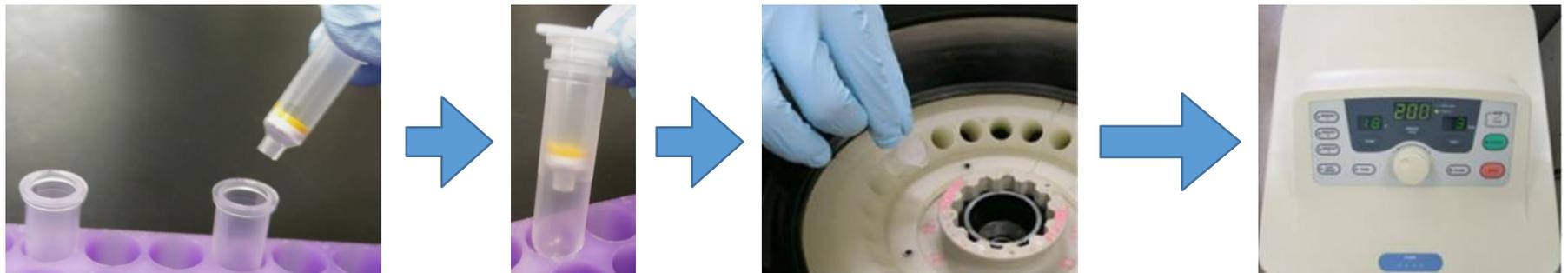


## [Extraction using QIAamp UCP DNA Micro Kit (3)]

6) Place the **QIAamp UCP MinElute Column** into a new collection tube and add **500 $\mu$ L** of **Buffer AW2**. Centrifuge at 20,000  $\times$  g (14,000 rpm) for 3 minutes.



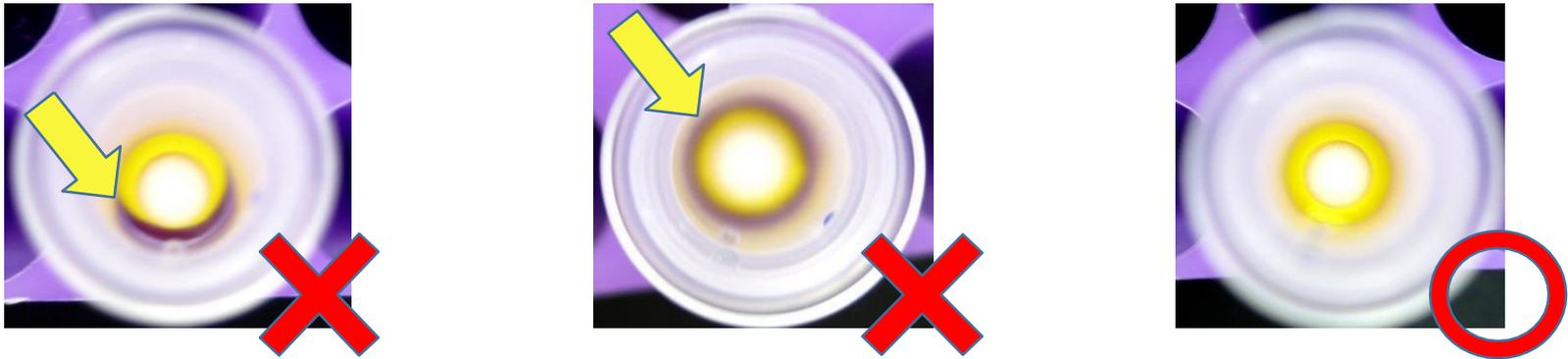
7) Place the **QIAamp UCP MinElute Column** into a new collection tube and centrifuge the empty **QIAamp UCP MinElute Column** at 20,000  $\times$  g (14,000 rpm) for 1 minute.



## [Extraction using QIAamp UCP DNA Micro Kit (4)]

### 8) Confirm that there is no remaining liquid in the **QIAamp UCP MinElute Column**

Note) The liquid in the example below is colored in blue for better visibility but actual liquid is transparent.



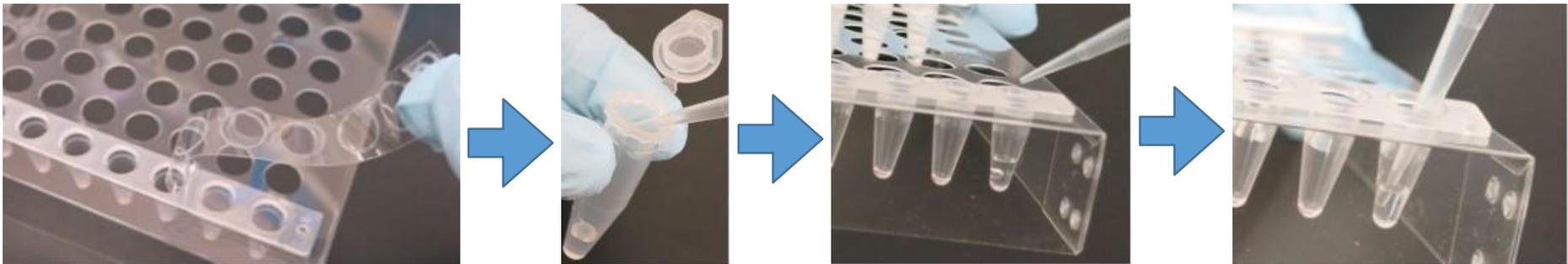
9) Place the **QIAamp UCP MinElute Column** into 1.5mL tube and add **100 $\mu$ L** of **Buffer AUE**. Incubate at room temperature (15 to 25°C) for 5 minutes. Centrifuge at 6,000  $\times$  g (8,000 rpm) for 1 minute and elute the sample.



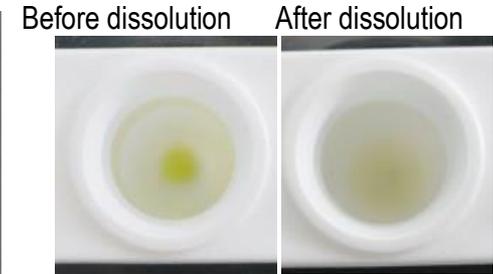
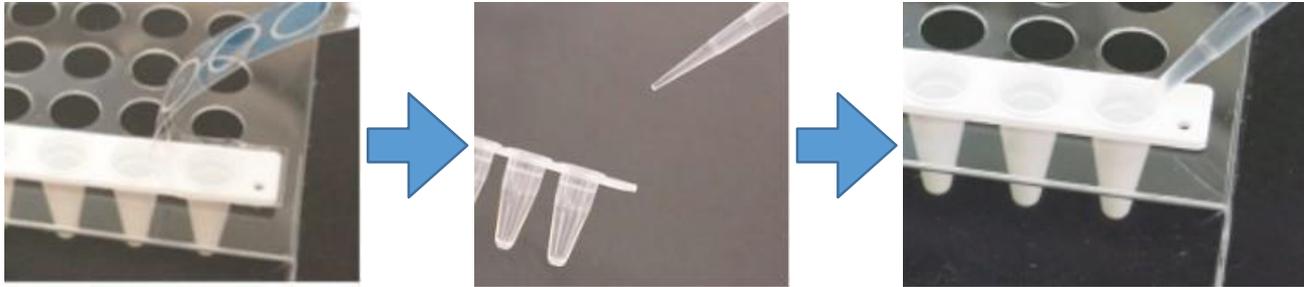
## Preparation of controls

- Preparation of positive control  
⇒ Add 1  $\mu$ L of positive control to 24  $\mu$ L of negative control.
- Preparation of negative control  
⇒ Use the 25  $\mu$ L of negative control as is.

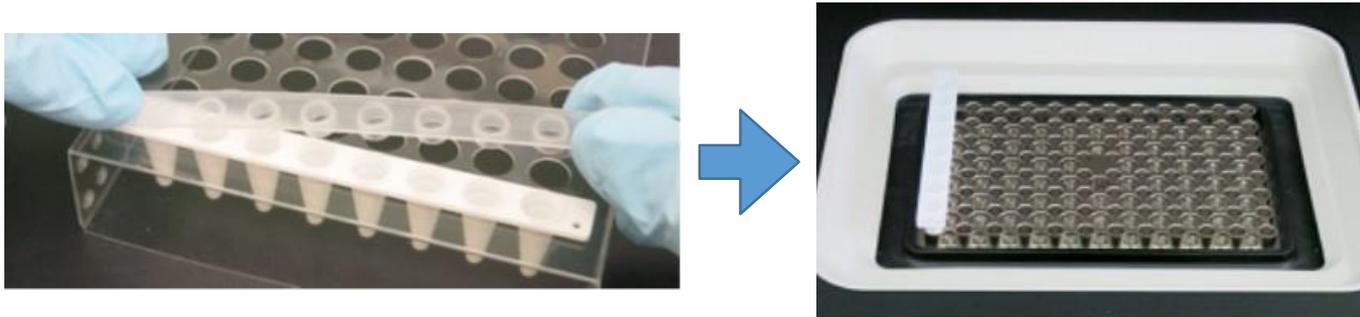
- 1) Take the necessary quantity of test strip A and test strip B out of the aluminum bag.
- 2) Peel off the film from test strip B and add 25  $\mu$ L each of the sample from DNA extraction, positive control, and negative control into respective wells, and pipet **gently** for 20 times to dissolve the solidified reagent. Dilute the sample as necessary before use.



3) Peel off the film from test strip A and move all the dissolved sample in test strip B into test strip A, and pipet **gently** for 20 times to dissolve the solidified reagent.

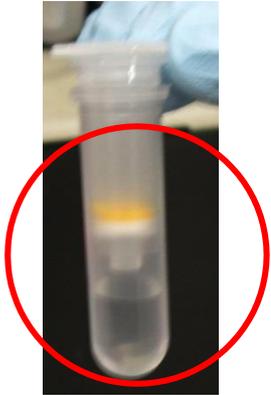


4) Attach flat cap to test strip A, and detect by real-time PCR.



\* When detection is performed using a transparent tube, first add the sample to test strip A, dissolve, and then move it into test strip B.

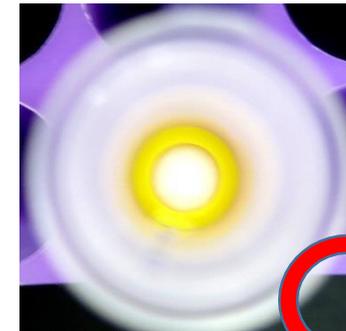
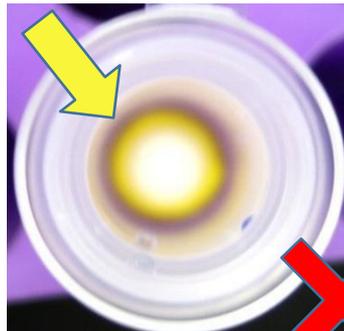
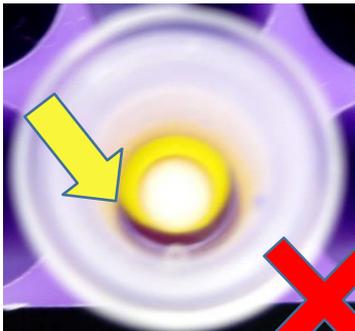
- 1) When performing the cleanup steps 4) through 7) of the extraction procedure using **QIAamp UCP MinElute Column**, be careful not to let the eluted sample in the collection tube contact the column.



Operate carefully so that the liquid in the collection tube will not touch the bottom end of the column.

- 2) When eluting a sample using **Buffer AUE**, any remaining **Buffer AUW2** in the column will interfere with PCR. Check for any remaining Buffer and if there is, remove it by pipetting, etc. without damaging the filter.

Note) The liquid in the picture below is colored in blue for better visibility but actual liquid is transparent.



The test strips after PCR have to be disposed of with the caps on and sealed in a bag, etc. following the rules of the relevant facility.

If PCR product scatters in the work environment, it can cause contamination or false-positive result.

**Make sure** to dispose of the test strips used for PCR with their caps on.

